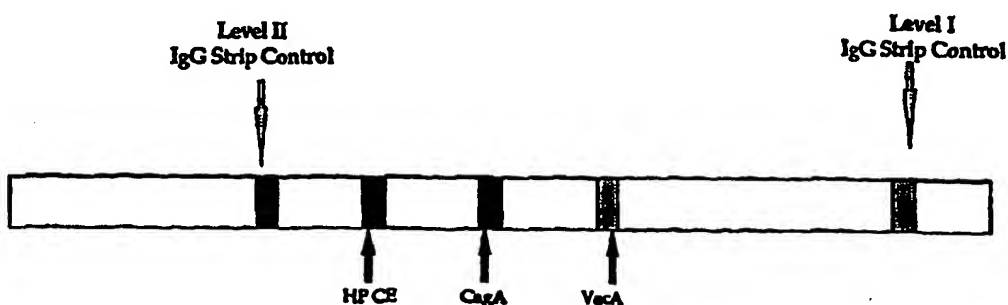




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(54) Title: *HELICOBACTER PYLORI* DIAGNOSTICS

(57) Abstract

Novel methods, membrane supports and immunodiagnostic test kits for diagnosing *Helicobacter pylori* infection, are disclosed. The methods can also be used to monitor the progress of treatment of an infection. The methods, supports and kits employ both type-common and type-specific *H. pylori* antigens and can conveniently be performed in a single-step assay format. The methods provide for highly accurate results and discriminate between *H. pylori* Type I and *H. pylori* Type II infection so that an accurate diagnosis can be accomplished.

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HELICOBACTER PYLORI DIAGNOSTICSBackground of the InventionTechnical Field

10 The present invention pertains generally to
bacterial diagnostic techniques. In particular, the
invention relates to methods for accurately detecting
Helicobacter pylori infection in a biological sample
and for monitoring the course of antibiotic treatment
15 in a patient with an *H. pylori* infection.

Background of the Invention

Helicobacter pylori, originally named
Campylobacter pylori, is a curved, microaerophilic,
20 gram-negative bacterium that exhibits high urease and
catalase activity. Recent studies suggest that *H.*
pylori infection may be either a cause of, or a
cofactor in, type B gastritis, peptic ulcers, and
gastric tumors. See, e.g., Blaser, *Gastroenterology*
25 (1987) 93:371-383; Dooley et al., *New Eng. J. Med.*
(1989) 321:1562-1566; Personnet et al., *New Eng. J.*
Med. (1991) 325:1127-1131. In this regard, *H. pylori*
colonizes the human gastric mucosa and causes an
infection that can persist for decades. Many people
30 with this condition are asymptomatic but are
nonetheless at a considerable risk of developing
peptic ulcers and/or gastric adenocarcinomas. For a
review of *H. pylori* and its role in gastric disease,
see, Telford et al., *Trends in Biotech.* (1994) 12:420-
35 426 and Blaser, M.J., *Scientific American* (February
1996):104-107.

H. pylori bacteria are divided into two groups, Type I and Type II, based on the presence or absence of specific proteins. In this regard, *H. pylori* produces several factors that function to establish and maintain infection. For example, both Type I and Type II bacteria include flagella that aid in mobility in the viscous mucus layer of the stomach. Both types of bacteria also produce ureases, presumably to neutralize the acid environment of the stomach. Additionally, the two types of bacteria produce a number of adhesins for tissue-specific colonization. On the other hand, only *H. pylori* Type I strains produce a potent cytotoxin, known as VacA or CT, as well as a surface-exposed immunodominant antigen which is associated with cytotoxin expression, known as CagA, CAI antigen or tagA. For descriptions of VacA and CagA, see, e.g., International Publication No. WO 93/18150, published 16 September 1993.

Patients with duodenal ulcers have been shown to produce antibodies to VacA and CagA and antibody titers appear to correlate with the severity of the disease. For example, in one study, more than 95% of patients with duodenal ulcer or duodenitis, and more than 70% of patients suffering from gastric ulcer, were found to be CagA seropositive. Telford et al., *Trends in Biotech.* (1994) 12:420-426. Furthermore, a correlation has been shown between CagA serum response and gastric adenocarcinoma. Telford et al., *supra*. Additionally, only cytotoxic strains are able to induce gastric lesions in a laboratory animal model. See, e.g., Telford et al., *J. Exp. Med.* (1994) 179:1653-1658. Thus, it is believed that only individuals infected with *H. pylori* Type I strains develop severe disease.

Several assays have been developed for the diagnosis of *H. pylori* infection. These assays,

unfortunately, suffer from several drawbacks. For example, bacterial culture assays have been described for the detection of *H. pylori*. U.S. Patent No. 5,498,528 describes such a method for detecting *H. pylori* in saliva. The assay requires incubating the test sample with a culture medium that supports the selective growth of *H. pylori*. The presence of the bacterium is detected by the activity of the enzyme urease which, as described above, is produced by *H. pylori*. Urease catalyzes the conversion of urea to ammonium causing an increase in the pH of the culture medium. The pH change can be detected by a color change to the medium due to the presence of a pH sensitive indicator. However, the assay is time consuming since the bacteria require a number of days for growth. The assay is also inconvenient and bacterial samples may degrade or become contaminated during transport to the laboratory.

Antibody detection tests provide an alternative to bacterial culture. In this regard, subjects colonized with *H. pylori* mount a humoral immune response and produce antibodies to the bacterium that can be used as a basis for diagnosis. IgA antibodies are found in gastric fluid while IgG antibodies are found in the circulation. However, such tests can suffer from a lack of specificity since *H. pylori* appears to be antigenically cross-reactive with *Campylobacter jejuni* and *C. coli*.

U.S. Patent No. 4,882,271 describes an *H. pylori* assay that utilizes high molecular weight cell-associated proteins, on the order of 300 kDa to 700 kDa, having urease activity, in an enzyme-linked immunosorbent assay (ELISA), in an attempt to circumvent the problems with cross-reactivity.

International Publication No. WO 96/12965, published 2 May 1996, describes an immunoblot assay

where a serological sample is reacted with two antigen components having molecular weights of 19.5 kDa, 26.5 kDa or 30 kDa, or alternatively, any one antigen component corresponding to a molecular weight of 35 kDa, 89 kDa, 116 kDa or 180 kDa. It is postulated by the inventors that the 19.5 kDa protein is a ferritin-like protein, the 26.5 and 30 kDa proteins are ureases, the 89 kDa protein is VacA, and that the 116 kDa protein is CagA. The 35 kDa and 180 kDa were uncharacterized.

Finally, European Patent Publication 329,570, published 23 August 1989, describes immunoassays for *H. pylori* infection using pooled suspensions of sonicates of several *H. pylori* strains, as well as immunoassays using purified *H. pylori* flagellae.

Although faster and more sensitive than bacterial culture, antibody detection tests, such as those described above, can give false positive and negative results and generally do not distinguish between *H. pylori* Type I and Type II infection. Thus, an additional test must be conducted to determine whether the infection is due to *H. pylori* Type I or Type II.

Accordingly, the wide spread availability of an accurate and efficient assay for *H. pylori* infection that readily distinguishes between Type I and Type II infection, would be important for the diagnosis of infection in both symptomatic and asymptomatic individuals.

Summary of the Invention

The present invention provides a simple, extremely accurate and efficient method for diagnosing *H. pylori* infection, as well as for distinguishing between *H. pylori* Type I and *H. pylori* Type II

infections. Thus, the method provides a technique for screening for individuals with *H. pylori* Type I infection. If Type I infection is detected, the individual can be given antibiotics to treat or
5 prevent type B gastritis, peptic ulcers, and gastric tumors. The method is also useful for monitoring the course of treatment in a patient with an *H. pylori* infection. The assay method utilizes both type-common antigens, as well as particular type-specific antigens
10 from the bacterium.

Accordingly, in one embodiment, the subject invention is directed to a method of detecting *H. pylori* infection comprising:

- (a) providing a biological sample;
- 15 (b) reacting the biological sample with one or more *H. pylori* type-common antigens and reacting the biological sample with one or more purified type-specific *H. pylori* Type I antigens, under conditions which allow *H. pylori* antibodies, when present in the
20 biological sample, to bind with the *H. pylori* type-common antigens and/or the type-specific antigens, thereby detecting the presence or absence of *H. pylori* infection.

In other embodiments, the invention is
25 directed to a method for distinguishing between *H. pylori* Type I and *H. pylori* Type II infection in a biological sample, or a method of monitoring a subject undergoing therapy for an *Helicobacter pylori* infection, the methods comprising:

- 30 (a) immobilizing one or more *H. pylori* type-common antigens, e.g., an *H. pylori* lysate and/or *H. pylori* urease, and immobilizing one or more purified type-specific *H. pylori* Type I antigens, e.g., *H. pylori* VacA and/or *H. pylori* CagA, on a nitrocellulose
35 strip;

(b) contacting the nitrocellulose strip from step (a) with the biological sample under conditions which allow anti-*H. pylori* Type I and anti-*H. pylori* Type II antibodies, when present in the biological sample, to bind with *H. pylori* type-common antigens present in the lysate and/or the type-specific *H. pylori* Type I antigens;

(c) removing unbound antibodies;

(d) providing a detectably labeled anti-human immunoglobulin antibody; and

(e) detecting the presence or absence of bound anti-human immunoglobulin antibodies in the biological sample,

thereby detecting the presence or absence of *H. pylori* Type I or Type II infection.

In particularly preferred embodiments, the biological sample is a human serum sample.

In yet further embodiments, the invention is directed to membrane supports comprising one or more *H. pylori* type-common antigens and one or more purified type-specific *H. pylori* Type I antigens, discretely immobilized thereon.

In another embodiment, the invention is directed to a nitrocellulose support comprising:

- (a) an *H. pylori* Type I VacA polypeptide;
- (b) an *H. pylori* Type I CagA polypeptide;
- (c) an *H. pylori* urease; and
- (d) a human IgG,

wherein the *H. pylori* polypeptides and urease, and the human IgG, are immobilized as discrete bands on said nitrocellulose support.

In a further embodiment, the invention is directed to a nitrocellulose support comprising:

- (a) an *H. pylori* Type I VacA polypeptide;
- (b) an *H. pylori* Type I CagA polypeptide;
- (c) an *H. pylori* lysate; and

(d) a human IgG,
wherein the *H. pylori* polypeptides and
lysate, and the human IgG, are immobilized as discrete
bands on said nitrocellulose support.

5 In other embodiments, the invention is
directed to immunodiagnostic test kits for detecting
H. pylori infection. The kits comprise (a) one or
more *H. pylori* type-common antigens; (b) one or more
purified type-specific *H. pylori* Type I antigens; and
10 (c) instructions for conducting the immunodiagnostic
test.

In still further embodiments, the invention
is directed to an immunodiagnostic test kit for
distinguishing between *H. pylori* Type I and *H. pylori*
15 Type II infection in a biological sample, or for
monitoring a subject undergoing therapy for an
Helicobacter pylori infection. The test kit comprises
(a) one or more *H. pylori* type-common antigens
immobilized on a nitrocellulose strip, e.g., an *H.*
20 *pylori* lysate and/or *H. pylori* urease; (b) one or more
purified type-specific *H. pylori* Type I antigens,
e.g., *H. pylori* VacA and/or *H. pylori* CagA,
immobilized on a nitrocellulose strip; and (c)
instructions for conducting the immunodiagnostic test.

25 These and other embodiments of the present
invention will readily occur to those of ordinary
skill in the art in view of the disclosure herein.

Brief Description of the Figures

30 Figure 1 depicts a representative test strip
for use in a strip immunoblot assay (SIA). Human IgG
is used as an internal control at two different levels
(Level I, low control; and Level II, high control).
CagA and VacA are used as the type-specific *H. pylori*
35 Type I antigens and HP CE denotes the *H. pylori* lysate
which contains type-common antigens.

Figure 2 shows another representative test strip for use in an SIA. As above, human IgG is used as an internal control at two different levels (Level I, low control; and Level II, high control). CagA and VacA are used as the type-specific *H. pylori* Type I antigens and are also used at two different levels to enhance the sensitivity of the assay as well as to monitor response to treatment. Urease is used as the type-common antigen.

Figures 3A-3B (SEQ ID NOS:____) show the nucleotide sequence and corresponding amino acid sequence for the *H. pylori* VacA antigen used in the SIAs described in the examples.

Figure 4 (SEQ ID NO:____) shows the nucleotide sequence and corresponding amino acid sequence for the *H. pylori* CagA antigen used in the SIAs described in the examples.

Detailed Description of the Invention

The practice of the present invention will employ, unless otherwise indicated, conventional methods of immunology, microbiology, molecular biology and recombinant DNA techniques within the skill of the art. Such techniques are explained fully in the literature. See, e.g., Sambrook, et al., *Molecular Cloning: A Laboratory Manual* (2nd Edition, 1989); *DNA Cloning: A Practical Approach*, vol. I & II (D. Glover, ed.); *Methods In Enzymology* (S. Colowick and N. Kaplan eds., Academic Press, Inc.); and *Handbook of Experimental Immunology*, Vols. I-IV (D.M. Weir and C.C. Blackwell eds., Blackwell Scientific Publications).

As used in this specification and the appended claims, the singular forms "a," "an" and "the" include plural references unless the content clearly dictates otherwise. Additionally, standard

abbreviations for nucleotides and amino acids are used in this specification.

I. Definitions

5 In describing the present invention, the following terms will be employed, and are intended to be defined as indicated below.

By "an *H. pylori* lysate" is meant an extract or lysate derived from an *H. pylori* Type I or Type II whole bacterium which includes one or more *H. pylori* polypeptides, as defined below, that reacts with antibodies generated against both of *H. pylori* Type I and *H. pylori* Type II. Such polypeptides are termed "type-common" antigens herein. Thus the term "lysate" as used herein refers to crude extracts that contain several *H. pylori* antigens, so long as at least one of the antigens present in the lysate is a type-common antigen. The lysate can be augmented with additional purified type-common and/or type-specific antigens. The term also denotes relatively purified compositions derived from such crude lysates which include only one or few such type-common antigens. Such lysates are prepared using techniques well known in the art, described further below.

25 Representative antigens that may be present in such lysates, either alone or in combination, include one or more type-common epitopes derived from the *H. pylori* adhesins such as, but not limited to, a 20 kDa N-acetyl-neuraminillactose-binding fibrillar haemagglutinin (HpaA), a 63 kDa protein that binds phosphatidylethanolamine and gangliotetraosyl ceramide, and a conserved fimbrial pilus-like structure. See, e.g., Telford et al., *Trends in Biotech.* (1994) 12:420-426 for a description of these antigens. Other type-common antigens that may be present in the lysate include one or more type-common

epitopes derived from any of the various flagellins such as the major flagellin, FlaA and the minor flagellin, FlaB. In this regard, the flagella of *H. pylori* are composed of FlaA and FlaB, each with a molecular weight of approximately 53 kDa. Either or both of FlaA and/or FlaB may be used as a source of type-common antigens for use with the present invention. Another representative type-common antigen includes *H. pylori* urease which is associated with the outer membrane and the periplasmic space of the bacterium. The holoenzyme is a large complex made up of two subunits of 26.5 kDa (UreA) and 61 kDa (UreB), respectively. Type-common epitopes derived from the holoenzyme, either of the subunits, or a combination of the three, can be present as the type-common antigen(s). Another representative type-common antigen that may be present in the lysate or used in further purified form includes the an *H. pylori* heat shock protein known as "hsp60." The DNA and corresponding amino acid sequences for hsp60 are known. See, e.g., International Publication No. WO 93/18150, published 16 September 1993. The full-length hsp60 antigen shown has about 546 amino acids and a molecular weight of about 58 kDa. It is to be understood that the lysate can also include other type-common antigens not specifically described herein.

By a "type-specific *H. pylori* Type I antigen" is meant a polypeptide, as defined below, derived from *H. pylori* Type I which reacts predominantly with antibodies against *H. pylori* Type I, but not with antibodies against *H. pylori* Type II. Representative type-specific *H. pylori* Type I antigens include: *H. pylori* VacA, also known as CT; and *H. pylori* CagA, also known as CAI antigen and tagA; and epitopes from these antigens which are capable of

reacting with antibodies against *H. pylori* Type I but not *H. pylori* Type II. Both VacA and CagA are discussed further below. It is to be understood that other type-specific *H. pylori* Type I antigens, not specifically described herein, are also captured by this definition.

The term "polypeptide" when used with reference to a type-common or type-specific *H. pylori* antigen, such as VacA, CagA or any of the other type-specific or type common antigens described above, refers to a VacA, CagA etc., whether native, recombinant or synthetic, which is derived from any of the various *H. pylori* strains. In the case of type-specific *H. pylori* Type I antigens, the polypeptide will be derived from an *H. pylori* Type I strain. In the case of a type-common antigen, the polypeptide may be derived from either of *H. pylori* Type I or Type II. The polypeptide need not include the full-length amino acid sequence of the reference molecule but can include only so much of the molecule as necessary in order for the polypeptide to react with the appropriate *H. pylori* antibodies. Thus, only one or few epitopes of the reference molecule need be present. Furthermore, the polypeptide may comprise a fusion protein between the full-length reference molecule or a fragment of the reference molecule, and another protein that does not disrupt the reactivity of the *H. pylori* polypeptide. It is readily apparent that the polypeptide may therefore comprise the full-length sequence, fragments, truncated and partial sequences, as well as analogs and precursor forms of the reference molecule. The term also intends deletions, additions and substitutions to the reference sequence, so long as the polypeptide retains the ability to react with *H. pylori* antibodies.

In this regard, particularly preferred substitutions will generally be conservative in nature, i.e., those substitutions that take place within a family of amino acids that are related in their side chains. Specifically, amino acids are generally divided into four families: (1) acidic -- aspartate and glutamate; (2) basic -- lysine, arginine, histidine; (3) non-polar -- alanine, valine, leucine, isoleucine, proline, phenylalanine, methionine, tryptophan; and (4) uncharged polar -- glycine, asparagine, glutamine, cystine, serine, threonine, tyrosine. Phenylalanine, tryptophan, and tyrosine are sometimes classified as aromatic amino acids. For example, it is reasonably predictable that an isolated replacement of leucine with isoleucine or valine, an aspartate with a glutamate, a threonine with a serine, or a similar conservative replacement of an amino acid with a structurally related amino acid, will not have a major effect on the biological activity. Proteins having substantially the same amino acid sequence as the reference molecule, but possessing minor amino acid substitutions that do not substantially affect the antibody binding capabilities of the protein, are therefore within the definition of the reference polypeptide.

By "VacA polypeptide" is meant a polypeptide as defined above which is derived from the antigen known as the *H. pylori* Type I Cytotoxin and which reacts predominantly with antibodies against *H. pylori* Type I, but not *H. pylori* Type II. The VacA protein induces vacuolization in epithelial cells in tissue culture and causes extensive tissue damage and ulceration when administered orally to mice. The DNA and corresponding amino acid sequences for VacA are known and reported in, e.g., International Publication No. WO 93/18150, published 16 September 1993. The

gene for the VacA polypeptide encodes a precursor of about 140 kDa that is processed to an active molecule of about 90-100 kDa. This molecule, in turn, is slowly proteolytically cleaved to generate two fragments that copurify with the intact 90 kDa molecule. See, Telford et al., *Trends in Biotech.* (1994) 12:420-426. Thus, the definition of "VacA polypeptide" as used herein includes the precursor protein, as well as the processed active molecule, proteolytic fragments thereof or portions or muteins thereof, which retain specific reactivity with antibodies present in a biological sample from an individual with *H. pylori* Type I infection. For example, the VacA polypeptide depicted in Figures 3A-3B and used in assays described herein includes a VacA fragment from Gly-311 to Ile-819, inclusive, of the full-length molecule, fused by a linker sequence of five amino acids to 154 amino acids of human SOD to facilitate recombinant expression.

By "CagA polypeptide" is meant a polypeptide as defined above which is derived from the *H. pylori* Type I cytotoxin associated immunodominant antigen and which reacts predominantly with antibodies against *H. pylori* Type I, but not *H. pylori* Type II. CagA is expressed on the bacterial surface. The DNA and corresponding amino acid sequences for CagA are known. See, e.g., International Publication No. WO 93/18150, published 16 September 1993. The full-length CagA antigen described therein includes about 1147 amino acids with a predicted molecular weight of about 128 kDa. The native protein shows interstrain size variability due to the presence of a variable number of repeats of a 102 bp DNA segment that encodes repeats of a proline-rich amino acid sequence. See, Covacci et al., *Proc. Natl. Acad. Sci. USA* (1993) 90:5791-5795. Accordingly, the reported molecular

weight of CagA ranges from about 120-135 kDa. Hence, the definition of "CagA polypeptide" as used herein includes any of the various CagA variants, fragments thereof and muteins thereof, which retain the ability to react with antibodies in a biological sample from an individual with *H. pylori* Type I infection but does not substantially react with antibodies generated against *H. pylori* Type II. For example, the CagA polypeptide depicted in Figure 4 and used in assays described herein is a truncated protein of 268 amino acids and includes Glu-748 to Glu-1015, inclusive, of the full-length molecule.

By "epitope" is meant a site on an antigen to which specific B cells and T cells respond. The term is also used interchangeably with "antigenic determinant" or "antigenic determinant site." An epitope can comprise 3 or more amino acids in a spatial conformation unique to the epitope. Generally, an epitope consists of at least 5 such amino acids and, more usually, consists of at least 8-10 such amino acids. Methods of determining spatial conformation of amino acids are known in the art and include, for example, x-ray crystallography and 2-dimensional nuclear magnetic resonance. Furthermore, the identification of epitopes in a given protein is readily accomplished using techniques well known in the art, such as by the use of hydrophobicity studies and by site-directed serology. See, also, Geysen et al., *Proc. Natl. Acad. Sci. USA* (1984) 81:3998-4002 (general method of rapidly synthesizing peptides to determine the location of immunogenic epitopes in a given antigen); U.S. Patent No. 4,708,871 (procedures for identifying and chemically synthesizing epitopes of antigens); and Geysen et al., *Molecular Immunology* (1986) 23:709-715 (technique for identifying peptides with high affinity for a given antibody). Antibodies

that recognize the same epitope can be identified in a simple immunoassay showing the ability of one antibody to block the binding of another antibody to a target antigen.

5 A "purified" protein or polypeptide is a protein which is recombinantly or synthetically produced, or isolated from its natural host, such that the amount of protein present in a composition is substantially higher than that present in a crude
10 preparation. In general, a purified protein will be at least about 50% homogeneous and more preferably at least about 80% to 90% homogeneous.

 As used herein, a "biological sample" refers to a sample of tissue or fluid isolated from an
15 individual, including but not limited to, for example, blood, plasma, serum, fecal matter, urine, bone marrow, bile, spinal fluid, lymph fluid, samples of the skin, external secretions of the skin, respiratory, intestinal, and genitourinary tracts,
20 samples derived from the gastric epithelium and gastric mucosa, tears, saliva, milk, blood cells, organs, biopsies and also samples of in vitro cell culture constituents including but not limited to conditioned media resulting from the growth of cells
25 and tissues in culture medium, e.g., recombinant cells, and cell components.

 As used herein, the terms "label" and "detectable label" refer to a molecule capable of detection, including, but not limited to, radioactive
30 isotopes, fluorescers, chemilumescers, enzymes, enzyme substrates, enzyme cofactors, enzyme inhibitors, chromophores, dyes, metal ions, metal sols, ligands (e.g., biotin or haptens) and the like. The term "fluorescer" refers to a substance or a
35 portion thereof which is capable of exhibiting fluorescence in the detectable range. Particular

examples of labels which may be used under the invention include fluorescein, rhodamine, dansyl, umbelliferone, Texas red, luminol, acradimum esters, NADPH and α - β -galactosidase.

5.

II. Modes of Carrying Out the Invention

The present invention is based on the discovery of novel diagnostic methods for accurately detecting *H. pylori* infection and for discriminating between *H. pylori* Type I and *H. pylori* Type II infection. The methods utilize one or more *H. pylori* type-common antigens, either purified or present in lysates derived from the bacterium, as well as purified type-specific *H. pylori* antigens. The use of both type-common and type-specific antigens reduces the incidence of false positive results. The methods can be practiced in a simple one-step assay format which allows for both detection of infection, as well as identification of the type of infection present, in a single assay. The method can also be practiced in two steps wherein the sample is first reacted with the *H. pylori* type-common antigens and if positive, reacted with one or more type-specific Type I antigens. The methods can also employ type-specific Type II antigens.

More particularly, the use of the *H. pylori* type-common antigens allows the diagnosis of *H. pylori* infection in general. The presence of one or more type-specific antigens allows determination of the bacterial type, i.e., whether the infection is caused by *H. pylori* Type I and/or *H. pylori* Type II. Due to the presence of the *H. pylori* type-common antigens, positive results will occur even in untypable samples. Hence, the incidence of false negatives is reduced. Furthermore, if *H. pylori* Type I infection is present, the individual can be administered antibiotics to

treat or prevent type B gastritis, peptic ulcers, and gastric tumors. Furthermore, the assays described herein are useful for monitoring the course of treatment in a subject to determine whether antibiotic therapy is effective.

The antigens for use in the subject diagnostic techniques can be produced using a variety of techniques.

For example, the type-common antigens can be provided in a lysate that can be obtained using methods well known in the art. Generally, such methods entail extracting type-common proteins from either *H. pylori* Type I or Type II bacteria using sonication, pressure disintegration, detergent extraction, fractionation, and the like. Type-common antigens present in such lysates can be further purified if desired, using standard purification techniques. *H. pylori* strains for use in such methods are readily available from several sources including the American Type Culture Collection (ATCC, Rockville, MD.). For example, ATCC strain designations NCTC 11637, 11639 and 11916, will find use as a source of the lysate. Other useful strains are known in the art.

The type-specific *H. pylori* Type I antigens can also be obtained using standard purification techniques. In this regard, particular antigens can be isolated from Type I *H. pylori* ulcer-producing strains using standard purification techniques such as column chromatography, electrophoresis, HPLC, immunoabsorbent techniques, affinity chromatography and immunoprecipitation. See, e.g., International Publication No. WO 96/12965, published 2 May 1996, for a description of the purification of several antigens from *H. pylori*. For example, ATCC strain designation NCTC 11916 is a Type I ulcer-producing strain of *H.*

pylori and can therefore be used as a source for one or more type-specific antigens for use in the subject invention.

5 The *H. pylori* antigens can also be generated using recombinant methods, well known in the art. In this regard, oligonucleotide probes can be devised based on the known sequences of the *H. pylori* genome and used to probe genomic or cDNA libraries for *H. pylori* genes encoding for the antigens useful in the present invention. The genes can then be further
10 isolated using standard techniques and, if desired, restriction enzymes employed to mutate the gene at desired portions of the full-length sequence.

Similarly, *H. pylori* genes can be isolated
15 directly from bacterial cells using known techniques, such as phenol extraction, and the sequence can be further manipulated to produce any desired alterations. See, e.g., Sambrook et al., *supra*, for a description of techniques used to obtain and isolate DNA. Finally, the genes encoding the *H. pylori*
20 antigens can be produced synthetically, based on the known sequences. The nucleotide sequence can be designed with the appropriate codons for the particular amino acid sequence desired. In general,
25 one will select preferred codons for the intended host in which the sequence will be expressed. The complete sequence is generally assembled from overlapping oligonucleotides prepared by standard methods and assembled into a complete coding sequence. See, e.g.,
30 Edge, *Nature* (1981) 292:756; Nambair et al., *Science* (1984) 223:1299; Jay et al., *J. Biol. Chem.* (1984) 259:6311.

Once coding sequences for the desired polypeptides have been isolated or synthesized, they
35 can be cloned into any suitable vector or replicon for expression in a variety of systems, including insect,

mammalian, bacterial, viral and yeast expression systems, all well known in the art. In particular, host cells are transformed with expression vectors which include control sequences operably linked to the
5 desired coding sequence.

The control sequences will be compatible with the particular host cell used. For example, typical promoters for mammalian cell expression include the SV40 early promoter, mouse mammary tumor
10 virus LTR promoter, adenovirus major late promoter (Ad MLP), and herpes simplex virus promoter, among others. Other non-viral promoters, such as a promoter derived from the murine metallothionein gene, will also find use in mammalian constructs. Mammalian expression may
15 be either constitutive or regulated (inducible), depending on the promoter. Typically, transcription termination and polyadenylation sequences will also be present, located 3' to the translation stop codon. Examples of transcription terminator/polyadenylation
20 signals include those derived from SV40 (Sambrook et al., *supra*). Introns, containing splice donor and acceptor sites, may also be designed into the constructs of the present invention.

Enhancer elements can also be used in the
25 mammalian constructs to increase expression levels. Examples include the SV40 early gene enhancer (Dijkema et al., *EMBO J.* (1985) 4:761) and the enhancer/promoters derived from the long terminal repeat (LTR) of the Rous Sarcoma Virus (Gorman et al.,
30 *Proc. Natl. Acad. Sci. USA* (1982b) 79:6777) and human cytomegalovirus (Boshart et al., *Cell* (1985) 41:521). A leader sequence can also be present which includes a sequence encoding a signal peptide, to provide for the secretion of the foreign protein in mammalian cells.
35 Preferably, there are processing sites encoded between the leader fragment and the gene of interest such that

the leader sequence can be cleaved either *in vivo* or *in vitro*. The adenovirus tripartite leader is an example of a leader sequence that provides for secretion of a foreign protein in mammalian cells.

5 Once complete, the mammalian expression vectors can be used to transform any of several mammalian cells. Methods for introduction of heterologous polynucleotides into mammalian cells are known in the art and include dextran-mediated
10 transfection, calcium phosphate precipitation, polybrene mediated transfection, protoplast fusion, electroporation, encapsulation of the polynucleotide(s) in liposomes, and direct
 microinjection of the DNA into nuclei.

15 Mammalian cell lines available as hosts for expression are also known and include many immortalized cell lines available from the American Type Culture Collection (ATCC), including but not limited to, Chinese hamster ovary (CHO) cells, HeLa cells,
20 baby hamster kidney (BHK) cells, monkey kidney cells (COS), human hepatocellular carcinoma cells (e.g., Hep G2), as well as others.

 The constructs of the present invention can also be expressed in yeast. Control sequences for
25 yeast vectors are known in the art and include promoters such as alcohol dehydrogenase (ADH) (EP Publication No. 284,044), enolase, glucokinase, glucose-6-phosphate isomerase, glyceraldehyde-3-phosphate-dehydrogenase (GAP or GAPDH), hexokinase,
30 phosphofructokinase, 3-phosphoglycerate mutase, and pyruvate kinase (PyK) (EP Publication No. 329,203). The yeast *PHO5* gene, encoding acid phosphatase, also provides useful promoter sequences (Myanohara et al., *Proc. Natl. Acad. Sci. USA* (1983) 80:1). In addition,
35 synthetic promoters which do not occur in nature also function as yeast promoters. For example, upstream

activating sequences (UAS) of one yeast promoter may be joined with the transcription activation region of another yeast promoter, creating a synthetic hybrid promoter. Examples of such hybrid promoters include
5 the ADH regulatory sequence linked to the GAP transcription activation region (U.S. Patent Nos. 4,876,197 and 4,880,734). Other examples of hybrid promoters include promoters which consist of the regulatory sequences of either the *ADH2*, *GAL4*, *GAL10*,
10 or *PHO5* genes, combined with the transcriptional activation region of a glycolytic enzyme gene such as GAP or PyK (EP Publication No. 164,556). Furthermore, a yeast promoter can include naturally occurring promoters of non-yeast origin that have the ability to
15 bind yeast RNA polymerase and initiate transcription.

Other control elements which may be included in the yeast expression vectors are terminators (e.g., from *GAPDH* and from the enolase gene (Holland, *J. Biol. Chem.* (1981) 256:1385), and leader sequences
20 which encode signal sequences for secretion. DNA encoding suitable signal sequences can be derived from genes for secreted yeast proteins, such as the yeast invertase gene (EP Publication No. 012,873; JPO Publication No. 62,096,086) and the α -factor gene
25 (U.S. Patent Nos. 4,588,684, 4,546,083 and 4,870,008; EP Publication No. 324,274; PCT Publication No. WO 89/02463). Alternatively, leaders of non-yeast origin, such as an interferon leader, also provide for secretion in yeast (EP Publication No. 060,057)

30 Expression and transformation vectors, either extrachromosomal replicons or integrating vectors, have been developed for transformation into many yeasts. For example, expression vectors have been developed for, *inter alia*, the following yeasts:
35 *Saccharomyces cerevisiae* (Hinnen et al., *Proc. Natl. Acad. Sci. USA* (1978) 75:1929; Ito et al., *J.*

- Bacteriol.* (1983) 153:163); *Saccharomyces carlsbergensis*; *Candida albicans* (Kurtz et al., *Mol. Cell. Biol.* (1986) 6:142); *Candida maltosa* (Kunze et al., *J. Basic Microbiol.* (1985) 25:141); *Hansenula polymorpha* (Gleeson et al., *J. Gen. Microbiol.* (1986) 132:3459; Roggenkamp et al., *Mol. Gen. Genet.* (1986) 202:302); *Kluyveromyces fragilis* (Das et al., *J. Bacteriol.* (1984) 158:1165); *Kluyveromyces lactis* (De Louvencourt et al., *J. Bacteriol.* (1983) 154:737; Van den Berg et al., *Bio/Technology* (1990) 8:135); *Pichia guilliermondii* (Kunze et al., *J. Basic Microbiol.* (1985) 25:141); *Pichia pastoris* (Cregg et al., *Mol. Cell. Biol.* (1985) 5:3376; U.S. Patent Nos. 4,837,148 and 4,929,555); *Schizosaccharomyces pombe* (Beach and Nurse, *Nature* (1981) 300:706); and *Yarrowia lipolytica* (Davidow et al., *Curr. Genet.* (1985) 10:380471; Gaillardin et al., *Curr. Genet.* (1985) 10:49).

Methods of introducing exogenous DNA into yeast hosts are well known in the art, and typically include either the transformation of spheroplasts or of intact yeast cells treated with alkali cations.

Bacterial expression systems can also be used with the present constructs. Control elements for use in bacteria include promoters, optionally containing operator sequences, and ribosome binding sites. Useful promoters include sequences derived from sugar metabolizing enzymes, such as galactose, lactose (*lac*) and maltose. Additional examples include promoter sequences derived from biosynthetic enzymes such as tryptophan (*trp*), the *b*-lactamase (*bla*) promoter system, bacteriophage lambda PL, and T5. In addition, synthetic promoters, such as the *tac* promoter (U.S. Patent No. 4,551,433), which do not occur in nature also function as in bacterial host cells.

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The foregoing systems are particularly compatible with *E. coli*. However, numerous other systems for use in bacterial hosts such as *Bacillus spp.*, *Streptococcus spp.*, and *Streptomyces spp.*, among others, are also known. Methods for introducing exogenous DNA into these hosts typically include the use of CaCl_2 or other agents, such as divalent cations and DMSO. DNA can also be introduced into bacterial cells by electroporation.

Other systems for expression of the desired antigens include insect cells and vectors suitable for use in these cells. The systems most commonly used are derived from the baculovirus *Autographa californica* polyhedrosis virus (AcNPV). Generally, the components of the expression system include a transfer vector, usually a bacterial plasmid, which contains both a fragment of the baculovirus genome, and a convenient restriction site for insertion of the heterologous gene or genes to be expressed; a wild type baculovirus with a sequence homologous to the baculovirus-specific fragment in the transfer vector (this allows for the homologous recombination of the heterologous gene into the baculovirus genome); and appropriate insect host cells and growth media.

Promoters for use in the vectors are typically derived from structural genes, abundantly transcribed at late times in a viral infection cycle. Examples include sequences derived from the gene encoding the viral polyhedron protein, Friesen et al., (1986) "The Regulation of Baculovirus Gene Expression" in: *The Molecular Biology of Baculoviruses* (ed. Walter Doerfler); EP Publication Nos. 127,839 and 155,476; and the gene encoding the p10 protein Vlak et al., *J. Gen. Virol.* (1988) 69:765. The plasmid usually also contains the polyhedrin polyadenylation signal (Miller et al., *Ann. Rev. Microbiol.* (1988) 42:177) and a

procaryotic ampicillin-resistance (*amp*) gene and origin of replication for selection and propagation in *E. coli*. DNA encoding suitable signal sequences can also be included and is generally derived from genes for secreted insect or baculovirus proteins, such as the baculovirus polyhedrin gene (Carbonell et al., *Gene* (1988) 73:409), as well as mammalian signal sequences such as those derived from genes encoding human α -interferon, Maeda et al., *Nature* (1985) 315:592; human gastrin-releasing peptide, Lebacqz-Verheyden et al., *Molec. Cell. Biol.* (1988) 8:3129; human IL-2, Smith et al., *Proc. Natl. Acad. Sci. USA* (1985) 82:8404; mouse IL-3, (Miyajima et al., *Gene* (1987) 58:273; and human glucocerebrosidase, Martin et al., *DNA* (1988) 7:99.

Currently, the most commonly used transfer vector for introducing foreign genes into AcNPV is pAc373. Many other vectors, known to those of skill in the art, have also been designed. These include, for example, pVL985 (which alters the polyhedrin start codon from ATG to ATT, and which introduces a *Bam*HI cloning site 32 bps downstream from the ATT; see Luckow and Summers, *Virology* (1989) 17:31).

The desired DNA sequence is inserted into the transfer vector, using known techniques (see, Summers and Smith, *supra*; Smith et al., *Mol. Cell. Biol.* (1983) 3:2156; and Luckow and Summers (1989) and

an insect cell host is cotransformed with the heterologous DNA of the transfer vector and the genomic DNA of wild type baculovirus--usually by cotransfection. The vector and viral genome are allowed to recombine. The packaged recombinant virus is expressed and recombinant plaques are identified and purified. Materials and methods for baculovirus/insect cell expression systems are

commercially available in kit form from, for example, Invitrogen, San Diego CA ("MaxBac" kit). These techniques are generally known to those skilled in the art and fully described in Summers and Smith, Texas Agricultural Experiment Station Bulletin No. 1555 (1987) (hereinafter "Summers and Smith").

Recombinant baculovirus expression vectors have been developed for infection into several insect cells. For example, recombinant baculoviruses have been developed for, *inter alia*: *Aedes aegypti*, *Autographa californica*, *Bombyx mori*, *Drosophila melanogaster*, *Spodoptera frugiperda*, and *Trichoplusia ni*.

It is often desirable that the polypeptides prepared using the above systems be fusion polypeptides. As with nonfusion proteins, these proteins may be expressed intracellularly or may be secreted from the cell into the growth medium.

Furthermore, plasmids can be constructed which include a chimeric gene sequence, encoding e.g., multiple type-specific *H. pylori* Type I antigens or multiple type-common antigens. The gene sequences can be present in a dicistronic gene configuration. Additional control elements can be situated between the various genes for efficient translation of RNA from the distal coding region. Alternatively, a chimeric transcription unit having a single open reading frame encoding the multiple antigens can also be constructed. Either a fusion can be made to allow for the synthesis of a chimeric protein or alternatively, protein processing signals can be engineered to provide cleavage by a protease, thus allowing liberation of the two or more proteins derived from translation of the template RNA. The processing protease may also be expressed in this

system either independently or as part of a chimera with the antigen coding region(s).

Viral systems, such as a vaccinia based infection/transfection system, as described in Tomei et al., *J. Virol.* (1993) 67:4017-4026 and Selby et al., *J. Gen. Virol.* (1993) 74:1103-1113, will also find use with the present invention. In this system, cells are first transfected *in vitro* with a vaccinia virus recombinant that encodes the bacteriophage T7 RNA polymerase. This polymerase displays exquisite specificity in that it only transcribes templates bearing T7 promoters. Following infection, cells are transfected with the DNA of interest, driven by a T7 promoter. The polymerase expressed in the cytoplasm from the vaccinia virus recombinant transcribes the transfected DNA into RNA which is then translated into protein by the host translational machinery. The method provides for high level, transient, cytoplasmic production of large quantities of RNA and its translation product(s).

Depending on the expression system and host selected, the antigens of the present invention are produced by growing host cells transformed by an expression vector under conditions whereby the antigen of interest is expressed. The antigen is then isolated from the host cells and purified. If the expression system provides for secretion of the antigen, the antigen can be purified directly from the media. If the antigen is not secreted, it is isolated from cell lysates. The selection of the appropriate growth conditions and recovery methods are within the skill of the art.

The *H. pylori* antigens may also be produced by chemical synthesis such as by solid phase or solution peptide synthesis, using methods known to those skilled in the art. Chemical synthesis of

peptides may be preferable if the antigen in question is relatively small. See, e.g., J. M. Stewart and J. D. Young, *Solid Phase Peptide Synthesis*, 2nd Ed., Pierce Chemical Co., Rockford, IL (1984) and G. Barany and R. B. Merrifield, *The Peptides: Analysis, Synthesis, Biology*, editors E. Gross and J. Meienhofer, Vol. 2, Academic Press, New York, (1980), pp. 3-254, for solid phase peptide synthesis techniques; and M. Bodansky, *Principles of Peptide Synthesis*, Springer-Verlag, Berlin (1984) and E. Gross and J. Meienhofer, Eds., *The Peptides: Analysis, Synthesis, Biology*, supra, Vol. 1, for classical solution synthesis.

The *H. pylori* type-common and type-specific *H. pylori* Type I antigens are used herein as diagnostics to detect the presence of reactive antibodies directed against the bacterium in a biological sample. Furthermore, the antigens can be used to monitor the course of antibiotic therapy by comparing results obtained at the outset of therapy to those obtained during and after a course of treatment. For example, the presence of antibodies reactive with the type-common and/or the type-specific *H. pylori* antigens can be detected using standard electrophoretic and immunodiagnostic techniques, including immunoassays such as competition, direct reaction, or sandwich type assays. Such assays include, but are not limited to, Western blots; agglutination tests; enzyme-labeled and mediated immunoassays, such as ELISAs; biotin/avidin type assays; radioimmunoassays; immunoelectrophoresis; immunoprecipitation, etc. The reactions generally include revealing labels such as fluorescent, chemiluminescent, radioactive, enzymatic labels or dye molecules, or other methods for detecting the

formation of a complex between the antigen and the antibody or antibodies reacted therewith.

5 The aforementioned assays generally involve separation of unbound antibody in a liquid phase from a solid phase support to which antigen-antibody complexes are bound. Solid supports which can be used in the practice of the invention include substrates such as nitrocellulose (e.g., in membrane or microtiter well form); polyvinylchloride (e.g., sheets or microtiter wells); polystyrene latex (e.g., beads or microtiter plates); polyvinylidene fluoride; diazotized paper; nylon membranes; activated beads, magnetically responsive beads, and the like.

15 Typically, a solid support is first reacted with a solid phase component (e.g., one or more type-common and/or one or more type-specific *H. pylori* Type I antigens) under suitable binding conditions such that the component is sufficiently immobilized to the support. Sometimes, immobilization of the antigen to the support can be enhanced by first coupling the antigen to a protein with better binding properties. Suitable coupling proteins include, but are not limited to, macromolecules such as serum albumins including bovine serum albumin (BSA), keyhole limpet hemocyanin, immunoglobulin molecules, thyroglobulin, ovalbumin, and other proteins well known to those skilled in the art. Other molecules that can be used to bind the antigens to the support include polysaccharides, polylactic acids, polyglycolic acids, polymeric amino acids, amino acid copolymers, and the like. Such molecules and methods of coupling these molecules to the antigens, are well known to those of ordinary skill in the art. See, e.g., Brinkley, M.A., *Bioconjugate Chem.* (1992) 3:2-13; Hashida et al., *J. Appl. Biochem.* (1984) 6:56-63; and Anjaneyulu and

Staros, *International J. of Peptide and Protein Res.*
(1987) 30:117-124.

After reacting the solid support with the
solid phase component, any non-immobilized solid-phase
5 components are removed from the support by washing,
and the support-bound component is then contacted with
a biological sample suspected of containing ligand
moieties (e.g., antibodies toward the immobilized
antigens) under suitable binding conditions. After
10 washing to remove any non-bound ligand, a secondary
binder moiety is added under suitable binding
conditions, where the secondary binder is capable of
associating selectively with the bound ligand. The
presence of the secondary binder can then be detected
15 using techniques well known in the art.

More particularly, an ELISA method can be
used, where the wells of a microtiter plate are coated
with the *H. pylori* type-common and/or the type-
specific *H. pylori* Type I antigen(s). A biological
20 sample containing or suspected of containing anti-*H.*
pylori immunoglobulin molecules is then added to the
coated wells. In assays where it is desired to use
one microtiter plate, a selected number of wells can
be coated with, e.g., a first type-specific antigen
25 moiety, a different set of wells coated with a second
type-specific antigen moiety and a third set of wells
with the *H. pylori* type-common antigen, etc. In the
alternative, a series of ELISAs can be run in tandem,
wherein individual plates are used for each type-
30 common and type-specific antigen moiety. After a
period of incubation sufficient to allow antibody
binding to the immobilized antigens, the plate(s) can
be washed to remove unbound moieties and a detectably
labeled secondary binding molecule added. The
35 secondary binding molecule is allowed to react with
any captured sample antibodies, the plate washed and

the presence of the secondary binding molecule detected using methods well known in the art.

Thus, in one particular embodiment, the presence of bound anti-type-specific and/or anti-*H. pylori* type-common antigen ligands from a biological sample can be readily detected using a secondary binder comprising an antibody directed against the antibody ligands. A number of anti-human immunoglobulin (Ig) molecules are known in the art (e.g., commercially available goat anti-human Ig or rabbit anti-human Ig). Ig molecules for use herein will preferably be of the IgG or IgA type, however, IgM may also be appropriate in some instances. The Ig molecules can be readily conjugated to a detectable enzyme label, such as horseradish peroxidase, glucose oxidase, Beta-galactosidase, alkaline phosphatase and urease, among others, using methods known to those of skill in the art. An appropriate enzyme substrate is then used to generate a detectable signal. In other related embodiments, competitive-type ELISA techniques can be practiced using methods known to those skilled in the art.

Assays can also be conducted in solution, such that the bacterial proteins and antibodies specific for those bacterial proteins form complexes under precipitating conditions. In one particular embodiment, the *H. pylori* type-common and/or the type-specific antigen(s) can be attached to a solid phase particle (e.g., an agarose bead or the like) using coupling techniques known in the art, such as by direct chemical or indirect coupling. The antigen-coated particle is then contacted under suitable binding conditions with a biological sample suspected of containing antibodies for *H. pylori* Type I or Type II. Cross-linking between bound antibodies causes the formation of particle-antigen-antibody complex

aggregates which can be precipitated and separated from the sample using washing and/or centrifugation. The reaction mixture can be analyzed to determine the presence or absence of antibody-antigen complexes using any of a number of standard methods, such as those immunodiagnostic methods described above.

In yet a further embodiment, an immunoaffinity matrix can be provided, wherein a polyclonal population of antibodies from a biological sample suspected of containing anti-*H. pylori* antibodies is immobilized to a substrate. In this regard, an initial affinity purification of the sample can be carried out using immobilized antigens. The resultant sample preparation will thus only contain anti-*H. pylori* moieties, avoiding potential nonspecific binding properties in the affinity support. A number of methods of immobilizing immunoglobulins (either intact or in specific fragments) at high yield and having good retention of antigen binding activity, are known in the art. Not being limited by any particular method, immobilized protein A or protein G can be used to immobilize immunoglobulins.

Accordingly, once the immunoglobulin molecules have been immobilized to provide an immunoaffinity matrix, the *H. pylori* type-common and type-specific antigens, having separate and distinct labels, are contacted with the bound antibodies under suitable binding conditions. After any non-specifically bound antigen has been washed from the immunoaffinity support, the presence of bound antigen can be determined by assaying for each specific label using methods known in the art.

A particularly preferred method for diagnosing *H. pylori* infection and monitoring the course of treatment using the present invention

involves the use of strip immunoblot assay (SIA) techniques, such as those known in the art which combine traditional Western and dot blotting techniques, e.g., the RIBA® (Chiron Corp., Emeryville, CA) test. In these assays, the *H. pylori* type-common and type-specific *H. pylori* Type I antigens are immobilized as individual, discrete portions, e.g., as bands or dots, on a membranous support. Thus, by "discretely immobilized" on a membrane support is meant that the antigens are present as separate components and are not mixed, such that reactivity or lack thereof with each of the antigens present can be assessed. A biological sample suspected of containing antibodies to *H. pylori* antigens is then reacted with the test membrane. Visualization of anti-*H. pylori* reactivity in the biological sample is accomplished using anti-human IgG enzyme-conjugates in conjunction with a colorimetric enzyme substrate. Internal controls, such as human IgM and human IgG, can also be present on the strip. The assay can be performed manually or used in an automated format.

Generally, the type-specific antigens, such as VacA and CagA, are applied to the strip in a concentration of about .5 to about 5 $\mu\text{g/ml}$, more preferably about .5 to 3 $\mu\text{g/ml}$ and most preferably about 1-2 $\mu\text{g/ml}$. Alternatively, two concentrations of antigen can be present, such as a low concentration and a high concentration, to provide a strip that can be used both for diagnosis of infection, as well as to monitor the response to treatment. Thus, for example, VacA and/or CagA can be provided in a concentration as specified above, as well as in one or more additional bands, in a concentration of about .005-.4 $\mu\text{g/ml}$, more preferably about .008-3 $\mu\text{g/ml}$ and most preferably about .1-.2 $\mu\text{g/ml}$.

The type-common antigen, e.g., the *H. pylori* lysate, can be applied at a concentration of about .25-2 $\mu\text{g/ml}$, more preferably about .25-1.5 $\mu\text{g/ml}$ and most preferably about .5-1 $\mu\text{g/ml}$. It is readily apparent that the concentration of antigen to be applied to the test strip will vary depending on the specific antigen used and can be readily determined by one of skill in the art.

The Ig controls, such as IgG, can be present in a single concentration, or in two concentrations, one low and one high. For example, IgG can be present in a concentration of about 50-250 ng/ml, more preferably about 75-200 ng/ml and most preferably about 100-185 ng/ml. A higher concentration of IgG can also be present along with the low concentration of IgG, to provide another internal control, such as at a concentration of about 400-1200 ng/ml, more preferably about 450-1000 ng/ml and most preferably about 500-950 ng/ml.

The above-described assay reagents, including the *H. pylori* lysate and type-specific antigens, optionally immobilized on a solid support, can be provided in kits, with suitable instructions and other necessary reagents, in order to conduct immunoassays as described above. The kit can also contain, depending on the particular immunoassay used, suitable labels and other packaged reagents and materials (i.e. wash buffers and the like). Standard immunoassays, such as those described above, can be conducted using these kits.

III. Experimental

Below are examples of specific embodiments for carrying out the present invention. The examples are offered for illustrative purposes only, and are

not intended to limit the scope of the present invention in any way.

Efforts have been made to ensure accuracy with respect to numbers used (e.g., amounts, temperatures, etc.), but some experimental error and deviation should, of course, be allowed for.

Example 1

Production of an *H. pylori* VacA Polypeptide

10 A VacA polypeptide was produced recombinantly as a fusion protein of 71.2 kDa which included 154 amino acids of human superoxide dismutase (SOD) (Hallewell et al., *Nucl. Acids Res.* (1985) 13:2017-2134), a linker of five amino acids (Asn-Leu-
15 Gly-Ile-Leu) and the VacA amino acid sequence Gly-311 through Ile-819 of *H. pylori* CCUG17874 (Covacci et al., *Proc. Natl. Acad. Sci. USA* (1993) 90:5791-5795; Telford et al., *J. Exp. Med.* (1994) 179:1653-1658).

In particular, DNA encoding for the
20 truncated VacA protein was synthesized by PCR and fused in frame with DNA sequences coding for SOD. The glucose regulated ADH2/GAPDH promoter (Cousens et al., *Gene* (1984) 61:265-275) was then incorporated at the 5'-end of the amplified fragment and the resulting
25 cassette was cloned into the yeast expression vector pBS24.1 (Pichuanes et al., *Protein Eng., Principle and Prac.* (1996) 5:129-161). This vector contains 2 μ and inverted repeat (IR) sequences for autonomous replication in yeast, the α -factor terminator to
30 ensure transcription termination, the *leu2-d* and *URA3* yeast genes for selection, and the β -lactamase gene and the ColE1 origin of replication for selection and propagation in *E. coli*. High expression levels of the *H. pylori* VacA recombinant protein were obtained in
35 *Saccharomyces cerevisiae* JSC310 (Mat a, *leu2*, *ura3-52*, *prb1-1122*, *pep4-3*, *prc1-407*, ::pDM15 (G418^R), [*cir*⁰]) as

evidenced by Coomassie-blue staining and immunoblot analysis of yeast proteins separated by SDS-PAGE. The nucleotide and corresponding amino acid sequence of the VacA recombinant protein is shown in Figures 3A-3B (SEQ ID NOS:_____).

The VacA recombinant protein was purified from yeast cells harvested several hours after depletion of glucose from the medium. This condition is needed to activate the ADH2/GAPDH promoter and trigger production of the foreign protein (Pichuantes et al., *J. Biol. Chem.* (1990) 265:13890-13898). Cells were broken with glass beads in a Dynomill using a lysis buffer containing 50mM Tris-HCl (pH 8.0), 150mM NaCl, 1mM EDTA, 1mM PMSF. The protein was recovered from the insoluble fraction (obtained by centrifugation at 48,400 x g for 30 minutes) with increasing amounts of urea (1M to 3M) in lysis buffer. After centrifugation at 48,400 x g for 30 minutes, the pellet containing the protein of interest was solubilized with lysis buffer containing 4M urea, 50mM DTT, 1N NaOH, while stirring on ice for 30 minutes. After removal of cell debris by centrifugation, the suspension was immediately titrated back to pH 8.0 with 6N NaOH. The supernatant was made 2.3% SDS, boiled for 3 minutes, cooled to room temperature and loaded onto a Sephacryl S-400 gel filtration column (Pharmacia) using a buffer containing PBS, 0.1mM EDTA, 0.1% SDS, pH 7.4. Fractions containing the recombinant VacA protein were pooled and concentrated in an Amicon concentrator (YM-10 membrane). After adjusting SDS to 2.3% and DTT to 50mM, the suspension was loaded back onto the same S-400 column for further fractionation. This procedure yielded a VacA protein >90 pure.

35

Example 2Production of an *H. pylori* CagA Polypeptide

A CagA polypeptide of 268 amino acids and having a molecular weight of 29.2 kDa, including amino acids Glu-748 through Glu-1015 of the CagA protein of the Chilean strain of *Helicobacter pylori*, Chetx-1, was produced recombinantly as follows. The DNA coding for this truncated CagA protein was synthesized by PCR and the initiation codon ATG was introduced at the 5'-end of the amplified fragment. The ADH2/GAPDH hybrid promoter (Cousens et al., Gene (1984) 61:265-275) was then incorporated at the 5'-end of the PCR-synthesized fragment and the resulting cassette was cloned into the yeast expression vector pBS24.1, essentially as described above. The resulting recombinant plasmid was used to transform *Saccharomyces cerevisiae* AD3 (Mat a, leu2, ura3-52, prb1-1122, pep4-3, prc1-407, ::pDM15 (G418^R), LEU2(Δ AD), [CIR⁰]) and expression of the recombinant antigen was monitored by Coomassie blue staining and immunoblot analysis of yeast proteins fractionated by SDS-PAGE. The nucleotide and corresponding amino acid sequence of the CagA recombinant protein is shown in Figure 4.

The CagA recombinant protein was purified from yeast cells suspended in lysis buffer (50mM Tris-HCl, 150mM NaCl, 1mM EDTA, 1mM PMSF, pH 8.0) and disrupted with glass beads in a Dynomill. After centrifugation at 48,400 x g for 30 minutes, the supernatant was made 4M urea, diluted 1/10 (v/v) with S Sepharose IEC equilibration buffer to OD₂₈₉ = 0.00 and the protein was eluted with a linear gradient of 0 to 0.5M NaCl in equilibration buffer. The elution peak containing the CagA protein was collected and precipitated with 30% ammonium sulfate. After centrifugation at 17,700 x g for 30 minutes, the pellet was discarded and the supernatant was

precipitated with a further 10% solid ammonium sulfate under the conditions described above. The pellet obtained after centrifugation at 17,700 x g for 30 minutes was dissolved in a minimum amount of PBS/8M urea, adjusted to 2.3% SDS/50mM DTT, boiled for 3 minutes, cooled to room temperature and loaded onto a Sephacryl S-300 gel filtration column (Pharmacia) using a buffer containing PBS, 0.1mM SDS, 1mM EDTA, pH 7.4. The fractions containing the protein of interest were collected and concentrated in an Amicon concentrator (YM-10 membrane). The concentrated material was made 2.3% SDS/50mM DTT and loaded back onto the same S-300 column for a second fractionation. This procedure yielded a CagA protein >90% pure.

15

Example 3

H. pylori Strip Immunoblot assay (SIA) Using Type-Specific and Type-Common H. pylori Antigens

A. An SIA was done as follows. An H. pylori extract containing a mixture of antigens including type-common antigens was obtained from Bioseed, Inc., Hillsborough, CA. Briefly, the extract was prepared from an H. pylori bacterium obtained from the American Type Culture Collection (ATCC) having an ATCC strain designation of ATCC 43504. The extract was prepared using detergent extraction and sonication.

The VacA and CagA type-specific antigens, described in Examples 1 and 2, respectively, were applied in discrete bands to nitrocellulose strips at concentrations of 1-2 µg/ml. The lysate including the type-common antigens was coated as another discrete band at a concentration of 0.5-1 µg/ml onto the same nitrocellulose strips. As internal controls, additional bands contained purified human IgG at a low (100-185 ng/ml) and high (500-925 ng/ml)

concentration. Other strips were coated with the antigens and lysate as described above, however, on these strips, the lysate was first enhanced with the VacA and CagA antigens added to the lysate at a concentration of .5-1 $\mu\text{g/ml}$ (VacA) and .5- $\mu\text{g/ml}$ (CagA). Figure 1 is a diagram of a nitrocellulose strip with the antigens as described above.

For the immunoblot assay, strips were processed in a batch fashion with 30 strips per batch. All steps were performed at room temperature. Each strip was numbered and then placed in a separate tube to which was added 1 ml of diluent (phosphate-buffered saline (PBS) with bovine protein stabilizers and detergents, 0.1% sodium azide and 0.05% gentamicin sulfate as preservatives) and 30 μl of a serum sample from an individual known to be infected with *H. pylori* Type I, suspected of being infected with *H. pylori* Type I or a control subject, was applied. The tubes were rocked gently for 4 h, the solution removed by aspiration, and 1 ml of fresh diluent was added to each tube. The tubes were rocked for 30 minutes, the solution removed by aspiration and 1 ml of wash buffer made from wash buffer concentrate (50X) (phosphate-buffered detergent solution with 0.01% thimerosal as a preservative) was added to each tube. The contents of each tube were emptied into a single wash vessel and the strips were washed by swirling for 20 seconds. The wash buffer was decanted and 30 ml of fresh buffer added and the process repeated. Residual solution was removed by aspiration and 20 ml of conjugate solution (peroxidase-labeled goat anti-human IgG (heavy and light chains), with bovine protein stabilizers, containing 0.01% thimerosal as a preservative) was added. The vessel was rotated at 110 rpm for 10 minutes, the conjugate solution was decanted and the wash step was repeated three times. Residual solution

was again removed by aspiration and 20 ml of substrate/developer (4-chloro-1-naphthol in methanol/phosphate-buffered hydrogen peroxide) added, followed by rotation for 15-20 minutes at 110 rpm.

5 The solution was decanted and the strips were washed twice in distilled water. Developed strips were placed face up on absorbent paper and allowed to dry for 30 minutes in the dark. Strips were then read within 3 h of drying. The intensity of the color of

10 the *H. pylori* antigen bands were assigned values ranging from - (0) to 4+ using the following algorithm. The low IgG band is assigned a value of 1+, the high IgG band is assigned a value of 3+. The lysate and VacA and CagA bands are then scored from 0

15 to 4+ according to how their band intensity compares to that of the IgG control bands.

Samples with any of the *H. pylori* antigen bands scoring 1+ or greater to the lysate band are considered to be positive for *H. pylori* infection.

20 Reactivity of 1+ or greater to the lysate band, in addition to reactivity of 1+ or greater to the CagA and/or VacA band, is considered positive for *H. pylori* Type I infection. Samples showing no reactivity of 1+ or greater to any bands are considered negative.

25 Using this assay, 29 samples obtained from individuals determined to be positive for *H. pylori* infection by endoscopy, 12 with duodenal ulcers, 9 with gastric ulcers and 8 with gastritis, were tested for reactivity. In another study, 80 individuals

30 known to have duodenal ulcers were tested. The results are shown in Tables 1 and 2, respectively. As can be seen, the assay is highly predictive of *H. pylori* infection.

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Table 1
Frequency of Band Reactivity
(*H. Pylori* Endoscopy Positive Specimens n=29)

	Lysate	Lysate + CagA	Lysate + CagA and/or VacA	CagA and/or VacA
D. Ulcer (n=12)	11/12 91.7%	10/12 83.3%	10/12 83.3%	1/12 8.3%
G. Ulcer (n=9)	9/9 100%	7/9 77.8%	8/9 88.9%	0/9 0.0%
Gastritis (n=8)	6/8 75.0%	4/8 50.0%	4/8 50.0%	1/8 12.5%

Table 2
Frequency of Band Reactivity in a
Duodenal Ulcer Population

Lysate	Lysate + CagA	Lysate + CagA and/or VacA	CagA and/or VacA
78/80 (97.5%)	71/80 (88.8%)	75/80 (93.8%)	2/80 (2.5%)

The above assay was also used to test samples from individuals undergoing antibiotic therapy for *H. pylori* infection. Table 3 shows a comparison between a person responding positively to antibiotic therapy (responder) and one not responding to therapy (non-responder). Samples from the responder were taken at 1, 3, 6, 9 and 12 months and samples from the non-responder taken at 8 and 16 months. As shown in Table 3, reactivity with the antigens decreased with time with the responder but did not significantly change with the non-responder.

Table 3 Therapy Monitoring of <i>H. pylori</i> Infections			
I. <u>Responder</u>			
	Lysate	CagA	VacA
Prebleed	3.42	1.92	0.31
1M	2.87	1.69	0.26
3M	2.32	1.09	0.33
6M	2.25	0.90	0.29
9M	2.13	0.73	0.21
12M	1.32	0.43	0.18
II. <u>Non-Responder</u>			
	Lysate	CagA	VacA
Prebleed	3.38	1.31	2.54
8M	3.06	0.99	2.49
16M	2.92	0.90	2.10

In another study, samples were obtained from individuals undergoing three different types of antibiotic regimens, as shown in Table 4. As can be seen, the response rate reported correlated with the results using the assay above for those individuals undergoing the triple and single antibiotic regimens.

Table 4 Therapy Monitoring of <i>H. pylori</i> Infection		
Therapy Regimen	Response Rate Reported	% Responders
Triple	73%	15/20 (75%)
Double	48%	11/17 (65%)
Single	19%	1/5 (20%)

Triple = Metronidazole + Bismuth + Amoxicillin

Double = Metronidazole + Bismuth

Single = Metronidazole

These two studies show that the assay is useful for determining whether an individual is responding to treatment.

B. Another SIA was done as follows. *H. pylori* urease was applied as a narrow band to nitrocellulose strips at a concentration of 1-2 $\mu\text{g/ml}$. The VacA antigen, described above, was applied at two concentrations, 1-2 $\mu\text{g/ml}$ and 0.1-0.2 $\mu\text{g/ml}$, to the same strips. The CagA type-specific antigen was also applied to the strips at the same two concentrations. As internal controls, additional bands contained purified human IgG at a low (100-185 ng/ml) and high (500-925 ng/ml) concentration. Figure 2 is a diagram of a nitrocellulose strip with the antigens as described above.

The assay was performed as described above.

The intensity of the color of the *H. pylori* antigen bands were assigned values, also as described above.

Thus, novel methods for detecting *H. pylori* infection as well as distinguishing between *H. pylori* Type I and Type II, and for monitoring the course of treatment, are disclosed. Although preferred embodiments of the subject invention have been described in some detail, it is understood that obvious variations can be made without departing from the spirit and the scope of the invention as defined by the appended claims.

CLAIMS

1. A method of detecting *Helicobacter pylori* infection in a subject comprising:
 - 5 (a) providing a biological sample from the subject; and
 - (b) reacting said biological sample with one or more *H. pylori* type-common antigens and reacting the biological sample with one or more purified type-specific *H. pylori* Type I antigens, under conditions
10 which allow *H. pylori* antibodies, when present in the biological sample, to bind with said type-common antigens and/or said type-specific antigen(s),
thereby detecting the presence or absence of
15 *H. pylori* infection in the subject.
2. The method of claim 1, wherein said *H. pylori* infection is *H. pylori* Type I.
- 20 3. The method of claim 1, wherein said *H. pylori* infection is *H. pylori* Type II.
4. The method of claim 1 further comprising:
 - 25 (c) removing unbound antibodies;
 - (d) providing one or more moieties capable of associating with said bound antibodies; and
 - (e) detecting the presence or absence of said one or more moieties,
30 thereby detecting the presence or absence of *H. pylori* infection.
5. The method of claim 4, wherein said one or more moieties comprises a detectably labeled anti-
35 human immunoglobulin antibody.

6. The method of claim 5 wherein the detectable label is a fluorescer or an enzyme.

5 7. The method of claim 1, wherein said one or more type-common antigens and said one or more type-specific antigens are immobilized on a solid support.

10 8. The method of claim 7 wherein said one or more type-common antigens and said one or more type-specific antigens are immobilized on the same solid support.

15 9. The method of claim 7 wherein said one or more type-common antigens and said one or more type-specific antigens are immobilized on different solid supports.

20 10. The method of claim 7 wherein the solid support is a nitrocellulose strip.

25 11. The method of claim 1, wherein said one or more type-specific antigens is selected from the group consisting of an *H. pylori* Type I VacA polypeptide and an *H. pylori* Type I CagA polypeptide.

30 12. The method of claim 1, wherein said one or more type-common antigens is provided in an *H. pylori* lysate.

13. The method of claim 1, wherein said one or more type-common antigens comprises an *H. pylori* urease.

35 14. The method of claim 1, wherein said biological sample is a human serum sample.

15. A method for distinguishing between *Helicobacter pylori* Type I and *Helicobacter pylori* Type II infection in a biological sample, said method
5 comprising:

- (a) immobilizing an *H. pylori* lysate comprising one or more *H. pylori* type-common antigens on a nitrocellulose strip, and immobilizing one or more purified type-specific *H. pylori* Type I antigens
10 on a nitrocellulose strip;
- (b) contacting said nitrocellulose strip from step (a) with said biological sample under conditions which allow anti-*H. pylori* Type I and anti-*H. pylori* Type II antibodies, when present in the
15 biological sample, to bind with *H. pylori* type-common antigens present in said lysate and/or said type-specific *H. pylori* Type I antigen(s);
- (c) removing unbound antibodies;
- (d) providing a detectably labeled anti-
20 human immunoglobulin antibody; and
- (e) detecting the presence or absence of bound anti-human immunoglobulin antibodies in said biological sample,
thereby detecting the presence or absence of
25 *H. pylori* Type I and/or *H. pylori* Type II infection.

16. The method of claim 15 wherein said one or more type-common antigens and said one or more type-specific antigens are immobilized on the same
30 nitrocellulose strip.

17. The method of claim 15, wherein said one or more type-common antigens and said one or more type-specific antigens are immobilized on different
35 nitrocellulose strips.

18. The method of claim 15, wherein said one or more type-specific antigens is selected from the group consisting of an *H. pylori* Type I VacA polypeptide and an *H. pylori* Type I CagA polypeptide.

5

19. The method of claim 15, wherein said biological sample is a human serum sample.

20. A method of monitoring a subject
10 undergoing therapy for an *Helicobacter pylori* infection comprising:
(a) providing a biological sample from the subject;
(b) immobilizing an *H. pylori* lysate
15 comprising one or more *H. pylori* type-common antigens on a nitrocellulose strip, and immobilizing one or more purified type-specific *H. pylori* Type I antigens on a nitrocellulose strip;
(c) contacting said nitrocellulose strip
20 from step (b) with said biological sample under conditions which allow anti-*H. pylori* Type I and anti-*H. pylori* Type II antibodies, when present in the biological sample, to bind with *H. pylori* type-common antigens present in said lysate and/or said type-specific
25 specific *H. pylori* Type I antigen(s);
(d) removing unbound antibodies;
(e) providing a detectably labeled anti-human immunoglobulin antibody; and
(f) detecting the presence or absence of
30 bound anti-human immunoglobulin antibodies in said biological sample,
thereby monitoring the course of treatment of the infection.

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21. The method of claim 20 wherein said one or more type-common antigens and said one or more type-specific antigens are immobilized on the same nitrocellulose strip.

5

22. The method of claim 20, wherein said one or more type-common antigens and said one or more type-specific antigens are immobilized on different nitrocellulose strips.

10

23. The method of claim 20, wherein said one or more type-specific antigens is selected from the group consisting of an *H. pylori* Type I VacA polypeptide and an *H. pylori* Type I CagA polypeptide.

15

24. The method of claim 20, wherein said biological sample is a human serum sample.

25. A membrane support comprising one or more *H. pylori* type-common antigens and one or more purified type-specific *H. pylori* Type I antigens, discretely immobilized thereon.

20

26. The membrane support of claim 25, wherein said one or more type-specific antigens is selected from the group consisting of an *H. pylori* Type I VacA polypeptide and an *H. pylori* Type I CagA polypeptide.

25

27. The membrane support of claim 25, wherein said one or more type-common antigens is provided in an *H. pylori* lysate.

30

28. The membrane support of claim 25, wherein said one or more type-common antigens comprises an *H. pylori* urease.

35

29. The membrane support of claim 25, further comprising a human immunoglobulin, discretely immobilized thereon.

5

30. The membrane support of claim 25, wherein the membrane support is a nitrocellulose strip.

10 31. A nitrocellulose support comprising:
(a) an *H. pylori* Type I VacA polypeptide;
(b) an *H. pylori* Type I CagA polypeptide;
(c) an *H. pylori* urease; and
(d) a human IgG,
15 wherein said *H. pylori* polypeptides and urease, and said human IgG, are immobilized as discrete bands on said nitrocellulose support.

20 32. A nitrocellulose support comprising:
(a) an *H. pylori* Type I VacA polypeptide;
(b) an *H. pylori* Type I CagA polypeptide;
(c) an *H. pylori* lysate; and
(d) a human IgG,
wherein said *H. pylori* polypeptides and
25 lysate, and said human IgG, are immobilized as discrete bands on said nitrocellulose support.

33. An immunodiagnostic test kit for detecting *H. pylori* infection, said test kit
30 comprising:
(a) one or more *H. pylori* type-common antigens; (b) one or more purified type-specific *H. pylori* Type I antigens; and
(c) instructions for conducting the
35 immunodiagnostic test.

34. The immunodiagnostic test kit of claim 33, wherein said one or more type-specific antigens is selected from the group consisting of an *H. pylori* Type I VacA polypeptide and an *H. pylori* Type I CagA polypeptide.

35. The immunodiagnostic test kit of claim 33, wherein said one or more type-common antigens are provided in an *H. pylori* lysate.

36. The immunodiagnostic test kit of claim 33, wherein said one or more type-common antigens comprises an *H. pylori* urease.

37. An immunodiagnostic test kit for distinguishing between *H. pylori* Type I and *H. pylori* Type II infection in a biological sample, said test kit comprising:

- (a) an *H. pylori* lysate comprising one or more *H. pylori* type-common antigens immobilized on a nitrocellulose strip;
- (b) one or more purified type-specific *H. pylori* Type I antigens immobilized on a nitrocellulose strip; and
- (c) instructions for conducting the immunodiagnostic test.

38. The immunodiagnostic test kit of claim 37, wherein said one or more type-specific antigens is selected from the group consisting of an *H. pylori* Type I VacA polypeptide and an *H. pylori* Type I CagA polypeptide.

39. The immunodiagnostic test kit of claim 37, wherein said lysate and said one or more type-

specific antigens are immobilized on the same nitrocellulose strip.

40. The immunodiagnostic test kit of claim 5 37, wherein said lysate and said one or more type-specific antigens are immobilized on different nitrocellulose strips.

41. An immunodiagnostic test kit for 10 monitoring a subject undergoing therapy for an *Helicobacter pylori* infection, said test kit comprising:

(a) an *H. pylori* lysate comprising one or more *H. pylori* type-common antigens immobilized on a 15 nitrocellulose strip;

(b) one or more purified type-specific *H. pylori* Type I antigens immobilized on a nitrocellulose strip; and

(c) instructions for conducting the 20 immunodiagnostic test.

42. The immunodiagnostic test kit of claim 41, wherein said one or more type-specific antigens is selected from the group consisting of an *H. pylori* 25 Type I VacA polypeptide and an *H. pylori* Type I CagA polypeptide.

43. The immunodiagnostic test kit of claim 41, wherein said lysate and said one or more type-specific antigens are immobilized on the same 30 nitrocellulose strip.

44. The immunodiagnostic test kit of claim 41, wherein said lysate and said one or more type-specific antigens are immobilized on different 35 nitrocellulose strips.

1/5

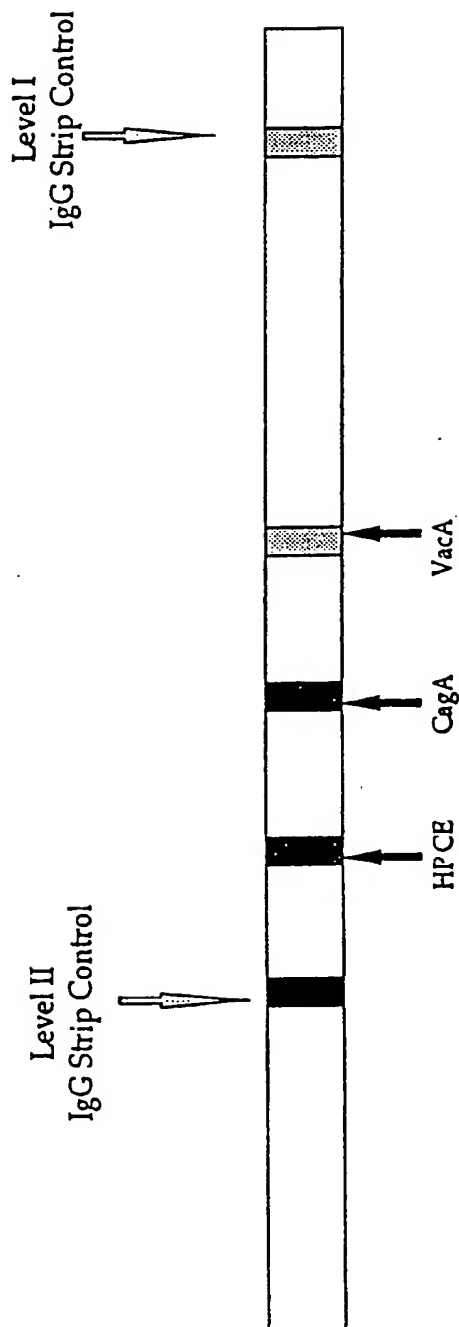


FIG. 1

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2/5

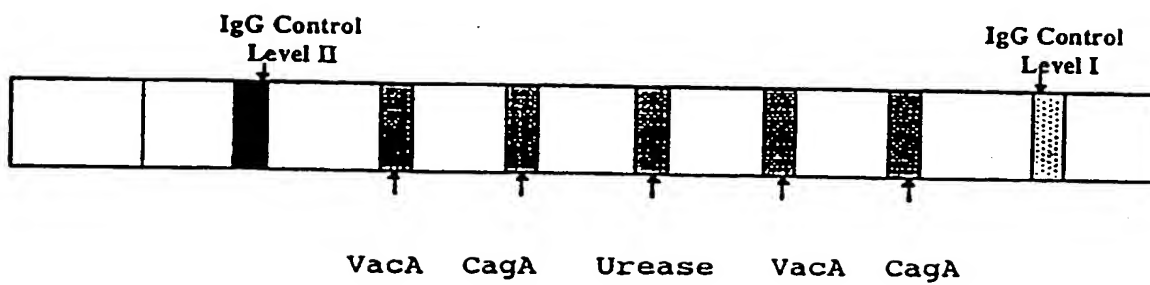


FIG. 2

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3/5

1 10
M A T K A V C V L K G D G P V Q G I I
CC ATG GCT ACA AAG GCT GTT TGT GTT TTG AAG GGT GAC GGC CCA GTT CAA GGT ATT ATT

20 30
N F E Q K E S N G P V K V W G S I K G L
AAC TTC GAG CAG AAG GAA AGT AAT GGA CCA GTG AAG GTG TGG GGA AGC ATT AAA GGA CTG

40 50
T E G L H G P H V H E P G D N T A G C T
ACT GAA GGC CTG CAT GGA TTC CAT GTT CAT GAG TTT GGA GAT AAT ACA GCA GGC TGT ACC

60 70
S A G P H F N P L S R K H G G P K D E E
AGT GCA GGT CCT CAC TTT AAT CCT CTA TCC AGA AAA CAC GGT GGG CCA AAG GAT GAA GAG

80 90
R H V G D L G N V T A D K D G V A D V S
AGG CAT GTT GGA GAC TTG GGC AAT GTG ACT GCT GAC AAA GAT GGT GTG GCC GAT GTG TCT

100 110
I E D S V I S L S G D H C I I G R T L V
ATT GAA GAT TCT GTG ATC TCA CTC TCA GGA GAC CAT TGC ATC ATT GGC CGC ACA CTG GTG

120 130
V H E K A D D L G K G G N E E S T K T G
GTC CAT GAA AAA GCA GAT GAC TTG GGC AAA GGT GGA AAT GAA GAA AGT ACA AAG ACA GGA

140 150
N A G S R L A C G V I G I A Q N L G I L
AAC GCT GGA AGT CGT TTG GCT TGT GGT GTA ATT GGG ATC GCC CAG AAT TTG GGA ATT CTC

160 170
G T L D L W Q S A G L N I I A P P E G G
GGC ACA CTG GAT TTG TGG CAA AGC GCC GGG TTA AAC ATT ATC GCT CCT CCA GAA GGT GGC

180 190
Y K D K P N N T P S Q S G A K N D K N E
TAT AAG GAT AAA CCC AAT AAT ACC CCT TCT CAA AGT GGT GCT AAA AAC GAC AAA AAT GAA

200 210
S A K N D K Q E S S Q N N S N T Q V I N
AGC GCT AAA AAC GAC AAA CAA GAG AGC AGT CAA AAT AAT AGT AAC ACT CAG GTC ATT AAC

220 230
P P N S A Q K T E V Q P T Q V I D G P F
CCA CCC AAT AGT GCG CAA AAA ACA GAA GTT CAA CCC ACG CAA GTC ATT GAT GGG CCT TTT

240 250
A G G K D T V V N I N R I N T N A D G T
GCG GGC GGC AAA GAC ACG GTT GTC AAT ATC AAC CGC ATC AAC ACT AAC GCT GAT GGC ACG

260 270
I R V G G F K A S L T T N A A R L H I G
ATT AGA GTG GGA GGG TTT AAA GCT TCT CTT ACC ACC AAT GCG GCT CAT TTG CAT ATC GGC

280 290
K G G V N L S N Q A S G R S L I V E N L
AAA GGC GGT GTC AAT CTG TCC AAT CAA GCG AGC GGG CGC TCT CTT ATA GTG GAA AAT CTA

300 310
T G N I T V D G P L R V N N Q V G G Y A
ACT GGG AAT ATC ACC GTT GAT GGG CCT TTA AGA GTG AAT AAT CAA GTG GCT GGC TAT GCT

320 330
L A G S S A N F E F K A G T D T K N G T
TTG GCA GGA TCA AGC GCG AAT TTT GAG TTT AAG GCT GGT ACG GAT ACC AAA AAC GGC ACA

340 350
A T F N N D I S L G R F V N L K V D A H
GCC ACT TTT AAT AAC GAT ATT AOT CTG GGA AGA TTT GTG AAT TTA AAG GTG GAT GCT CAT

FIG. 3A
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4/5

360 T A N F K G I D T G 370 G G F N T L D F S
ACA GCT AAT TTT AAA GGT ATT GAT ACG GGT AAT GGT GGT TTC AAC ACC TTA GAT TTT AGT

380 Q V T D K V N I N K L 390 I T A S T N V A V
GGC GTT ACA GAC AAA GTC AAT ATC AAC AAG CTC ATT ACG GCT TCC ACT AAT GTG GCC GTT

400 K N P N I N E L I V 410 K T N G I S V G E Y
AAA AAC TTC AAC ATT AAT GAA TTG ATT GTT AAA ACC AAT GGG ATA AGT GTG GGG GAA TAT

420 T H F S E D I G S Q 430 S R I N T V R L E T
ACT CAT TTT AGC GAA GAT ATA GGC AGT CAA TCG CGC ATC AAT ACC GTG CGT TTG GAA ACT

440 G T R S L F S G G V 450 K F K G G E K L V I
GGC ACT AGG TCA CTT TTC TCT GGG GGT GTT AAA TTT AAA GGT GGC GAA AAA TTG GTT ATA

460 D E P Y Y S P W N Y F 470 D A R N I K N V E
GAT GAG TTT TAC TAT AGC CCT TGG AAT TAT TTT GAC GCT AGA AAT ATT AAA AAT GTT GAA

480 I T N K L A F G P Q 490 G S P W G T S K L M
ATC ACC AAT AAA CTT GCT TTT GGA CCT CAA GGA AGT CCT TGG GGC ACA TCA AAA CTT ATG

500 F N N L T L Q Q N A V 510 M D Y S Q F S N L
TTC AAT AAT CTA ACC CTA GGT CAA AAT GCG GTC ATG GAT TAT AGC CAA TTT TCA AAT TTA

520 T I Q G D F I N N Q 530 G T I N Y L V R G G
ACC ATT CAA GGG GAT TTC ATC AAC AAT CAA GGC ACT ATC AAC TAT CTG GTC CGA GGT GGG

540 K V A T L S V G N A 550 A A M M P N N D I D
AAA GTG GCA ACC TTA AGC GTA GGC AAT GCA GCA GCT ATG ATG TTT AAT AAT GAT ATA GAC

560 S A T G F Y K P L I K 570 I N S A Q D L I K
AGC GCG ACC GGA TTT TAC AAA CCG CTC ATC AAG ATT AAC AGC GCT CAA GAT CTC ATT AAA

580 N T E H V L L K A K I 590 I I G Y G N V S T G
AAT ACA GAA CAT GTT TTA TTG AAA GCG AAA ATC ATT GGT TAT GGT AAT GTT TCT ACA GGT

600 T N G I S N V N L E 610 E Q F K E R L A L Y
ACC AAT GGC ATT AGT AAT GTT AAT CTA GAA GAG CAA TTC AAA GAG CGC CTA GCC CTT TAT

620 N N N N R M D T C V 630 V R N T D D I K A C
AAC AAC AAT AAC CGC ATG GAT ACT TGT GTG GTG CGA AAT ACT GAT GAC ATT AAA GCA TGC

640 G M A I G D Q S M V N 650 N N P D N Y K Y L I
GGT ATG GCT ATC GGC GAT CAA AGC ATG GTG AAC AAC CCT GAC AAT TAC AAG TAT CTT ATC

660 G K A W K N I G I OC
GGT AAG GCA TGG AAA AAT ATA GCG ATC TAA TAGGTCGAC

FIG. 3B

5/5

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1      10      20
M   E   F   K   N   G   K   N   K   D   F   S   K   V   T   Q   A   K   S   D
ATG GAA TTC AAA AAT GGC AAA AAT AAG GAT TTC AGC AAG GTA ACG CAA GCA AAA AGC GAC

      30      40
L   E   N   S   I   K   D   V   I   F   N   Q   K   I   T   D   K   V   D   D
CTT GAA AAT TCC ATT AAA GAT GTG ATT TTC AAT CAA AAG ATA ACG GAT AAA GTT GAT GAT

      50      60
L   N   Q   A   V   S   V   A   K   A   T   G   D   F   S   R   V   E   Q   A
CTC AAT CAA GCG GTA TCA GTG GCT AAA GCA ACG GGT GAT TTC AGT AGG GTA GAG CAA GCG

      70      80
L   A   D   L   K   N   F   S   K   E   Q   L   A   Q   Q   A   Q   K   N   E
TTA CCC GAT CTC AAA AAC TTC TCA AAG GAG CAA TTG GCT CAA CAA GCT CAA AAA AAT GAA

      90      100
S   L   N   A   G   K   K   S   E   I   Y   Q   S   V   K   N   G   V   N   G
AGT CTC AAT GCT GGA AAA AAA TCT GAA ATA TAC CAA TCC GTT AAG AAT GGT GTA AAC GGA

      110      120
T   L   V   G   N   G   L   S   Q   A   E   A   T   T   L   S   K   N   F   S
ACC CTA GTC GGT AAT GCG TTA TCT CAA GCA GAA GCC ACA ACT CTT TCT AAA AAC TTT TCG

      130      140
D   I   K   K   E   L   N   A   K   L   F   G   N   F   N   N   N   N   N   N
GAC ATC AAG AAA GAG TTG AAT CCA AAA CTT TTT GGA AAT TTC AAT AAC AAT AAC AAT AAT

      150      160
G   L   K   N   S   T   E   P   I   Y   A   K   V   N   K   K   K   T   G   Q
GGT CTC AAA AAC AGC ACA GAA CCC ATT TAT GCT AAA GTT AAT AAA AAG AAA ACA GGA CAA

      170      180
V   A   S   P   E   E   P   I   Y   T   Q   V   A   K   K   V   N   A   K   I
GTA GCT AGC CCT GAA GAA CCC ATT TAT ACT CAA GTT GCT AAA AAG GTA AAT GCA AAA ATT

      190      200
D   R   L   N   Q   I   A   S   G   L   G   G   V   G   K   A   A   G   F   P
GAC CGA CTC AAT CAA ATA GCA AGT GGT TTG GGT GGT GTA GGG AAA GCA GCG GCG TTC CCT

      210      220
L   K   R   H   D   K   V   D   D   L   S   K   V   G   R   S   V   S   P   E
TTG AAA AGG CAT GAT AAA GTT GAT GAT CTC AGT AAG GTA GGG CGA TCA GTT AGC CCT GAA

      230      240
P   I   Y   A   T   I   D   D   L   G   G   P   F   P   L   K   R   H   D   K
CCC ATT TAT GCT ACG ATT GAT GAT CTC GGC GGA CCT TTC CCT TTG AAA AGG CAT GAT AAA

      250      260
V   D   D   L   S   K   V   G   L   S   R   N   Q   E   L   A   Q   K   I   D
GTT GAT GAT CTC AGT AAG GTA GGG CTT TCA AGG AAC CAA GAA TTG GCT CAG AAA ATT GAC

      268
N   L   N   Q   A   V   S   E   OC
AAT CTC AAT CAA GCG GTA TCA GAA TAA TAGTCGAC

Translated Mol. Weight = 29195.84

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FIG. 4

SUBSTITUTE SHEET (RULE 26)

INTERNATIONAL SEARCH REPORT

International Application No

PCT/US 97/22798

A. CLASSIFICATION OF SUBJECT MATTER

IPC 6 G01N33/569 G01N33/543

According to International Patent Classification(IPC) or to both national classification and IPC

B. FIELDS SEARCHED

Minimum documentation searched (classification system followed by classification symbols)

IPC 6 G01N

Documentation searched other than minimum documentation to the extent that such documents are included in the fields searched

Electronic data base consulted during the international search (name of data base and, where practical, search terms used)

C. DOCUMENTS CONSIDERED TO BE RELEVANT

Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>WO 93 18150 A (BIOCINE SCLAVO S.P.A.) 16 September 1993 cited in the application see the whole document</p> <p style="text-align: center;">--- -/--</p>	1-44

☒ Further documents are listed in the continuation of box C.

☒ Patent family members are listed in annex.

* Special categories of cited documents :

"A" document defining the general state of the art which is not considered to be of particular relevance

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"P" document published prior to the international filing date but later than the priority date claimed

"T" later document published after the international filing date or priority date and not in conflict with the application but cited to understand the principle or theory underlying the invention

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"Y" document of particular relevance; the claimed invention cannot be considered to involve an inventive step when the document is combined with one or more other such documents, such combination being obvious to a person skilled in the art.

"&" document member of the same patent family

Date of the actual completion of the international search

23 April 1998

Date of mailing of the international search report

08/05/1998

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Van Bohemen, C

INTERNATIONAL SEARCH REPORT

Inter. Patent Application No
PCT/US 97/22798

C.(Continuation) DOCUMENTS CONSIDERED TO BE RELEVANT		
Category *	Citation of document, with indication, where appropriate, of the relevant passages	Relevant to claim No.
Y	<p>CHEMICAL ABSTRACTS, vol. 126, no. 1, 7 July 1997 Columbus, Ohio, US; abstract no. 747006, XP002063245 see abstract & S. CENSINI ET AL.: "CAG, a pathogenicity island of Helicobacter pylori, encodes types I-specific and disease-associated virulence factors " PROCEEDINGS OF THE NATIONAL ACADEMY OF SCIENCES USA, vol. 93, no. 25, 1 December 1996, WASHINGTON DC USA, pages 14648-14653, -----</p>	1-44

INTERNATIONAL SEARCH REPORT

information on patent family members

International Application No

PCT/US 97/22798

Patent document cited in search report	Publication date	Patent family member(s)	Publication date
WO 9318150 A	16-09-93	IT 1262895 B	22-07-96
		CA 2131729 A	16-09-93
		EP 0643770 A	22-03-95
		JP 7504565 T	25-05-95
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